

1 16S rRNA phylogeny of sponge-associated cyanobacteria

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ABSTRACT

1
2 Phylogenetic analyses of 16S rDNA sequences of sponge-associated
3 cyanobacteria showed them to be polyphyletic, implying that they derived from
4 multiple independent symbiotic events. Most of the symbiont sequences were
5 affiliated to a group of *Synechococcus* and *Prochlorococcus* species. However, other
6 symbionts were related to different groups, such as the Oscillatoriales.

1 Although both cyanobacteria and sponges have a very long evolutionary
2 history (1,3), little is known about the identity and phylogeny of cyanobacterial
3 sponge symbionts. For marine sponges symbiosis with cyanobacteria can be obligate
4 or non-obligate according to the sponge species (2). Thus far, all attempts of culturing
5 sponge-associated cyanobacteria have failed (unpublished data, 13). It is thus not
6 known whether sponge-associated cyanobacteria are able to survive outside their host.
7 The most common sponge-associated cyanobacterium was not found in water samples
8 (13, 15). In *Chondrilla australiensis* this cyanobacterium is transmitted vertically
9 (through sponge eggs) (14). Additionally, a study showing co-speciation between
10 *Dysidea* species and their associated cyanobacteria (12) supports the hypothesis that
11 sponges and associated cyanobacteria are coevolving. Few studies sequenced the 16S
12 rRNA gene of sponge-associated cyanobacteria: Diaz (5) identified the first two
13 cyanobacteria from marine sponges, Webb and Maas (16) found that the
14 cyanobacteria inhabiting *Mycale hentscheli* were phylogenetically related to
15 *Cyanobacterium stanieri* and species of *Synechocystis* and *Prochloron*, and Hentschel
16 *et al.* (7) found that 7 sequences from cyanobacteria inhabiting the sponges *Aplysina*
17 *aerophoba* and *Theonella swinhoei* could be divided into two clades
18 (*Synechococcus/Prochlorococcus* and *Pleurocapsa*). The aim of the present study was
19 to increase our understanding on the diversity of sponge-associated cyanobacteria, and
20 to determine their phylogenetic position.

21 Sponge samples from 16 species were collected from four locations: The
22 Caribbean (Bahamas, 26°33'N, 77°52'W), Mediterranean (Rapallo, Italy, 44°18'N,
23 9°12'E), Red Sea (Elat, 31°35'N, 34°54'E), and Western Indian Ocean (Zanzibar,
24 06°09'S, 39°11'E). Aposymbiotic specimens (specimens that do not contain
25 cyanobacterial symbionts), growing in dark caves or overhangs, were also collected

1 for two sponge species (*Petrosia ficiformis* and *Xestospongia muta*). Those samples,
2 collected at short distances from symbiotic specimens, were used as negative controls,
3 to ensure that 16S rDNA sequences were derived from true symbionts and not from
4 surface associated cyanobacteria or digested cyanobacteria. Photosynthetic activity
5 inside the living sponge tissue was tested by pulse amplitude modulated fluorometry
6 (Diving PAM, Walz, Germany). Tissue samples (1 cm³) were rinsed twice in 100%
7 EtOH, and kept in 100% EtOH at 4 °C. DNA was extracted following the procedure
8 of Bernatzky and Tanksley (4). 16S rDNA was amplified with the primers 361F (5'-
9 GAATTTTCCGCAATGGGC -3') and 1459R (5'- GGTAAYGACTTCGGGCRT -
10 3') (5). 1060-bp fragments were cloned in the PTZ57R/T vector (Fermentas). Twenty
11 clones per individual were amplified using M13 universal primers. The PCR products
12 were digested with restriction enzymes *ApaI* and *HaeIII*. One clone was sequenced for
13 each pattern present in more than 10% of the clones. The sequences were deposited in
14 Genbank (accession numbers AY701287-AY701315).

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16 Neither photosynthetic activity, nor amplification of the 16S rRNA gene was
17 obtained for aposymbiotic specimens. On the opposite, photosynthetic activity was
18 recorded for all the other samples. Up to three different cyanobacterial clones were
19 sequenced per individual sponge investigated (with a sequence homology of 90-
20 99.7%). Similarly, Webb and Maas (16) showed four closely related (99.1-99.8%)
21 cyanobacterial clones in the sponge *Mycale hentscheli*. Different cyanobacterial types
22 from the same individual were usually phylogenetically close to each other (<1-2%
23 sequence divergence), but in one case (*Lendenfeldia dendyi*) the symbionts were very
24 divergent: two types were in group 5, while the third type was in group 2, with 10%
25 sequence divergence from the former two (Fig. 1). Closely related cyanobacterial

1 types should not be the result of Taq-polymerase errors or cloning-bias since only
2 patterns present in more than one clone were sequenced. However, the true
3 cyanobacterial diversity in sponges might be underestimated. Nevertheless our results
4 are in agreement with other studies (13, 16). For example, Usher et al. (13) also
5 identified only one cyanobacterial type in the sponge *Petrosia ficiformis*.

6 A maximum likelihood (ML, (6)) analysis was done based on 147 taxa and
7 1396 nucleotides. The sequences were chosen to include: 1- representatives of the
8 cyanobacteria diversity; 2- all sponge-associated cyanobacteria sequences overlapping
9 with the new 16S rDNA sequences; and 3- representatives of the *Synechococcus* and
10 *Prochlorococcus* diversity because sponge symbionts have been suggested to belong
11 to these genera. BLAST searches were also conducted for each sponge sequence in
12 order to include in the analysis the most similar sequences available in the databanks.
13 The GTR model with a gamma distribution (4 categories), and a proportion of
14 invariant site was found to be the most appropriate using Modeltest version 3.06 (9)).
15 The ML tree was reconstructed in an iterative way using PAUP* version 4.0b10 (11).
16 First a heuristic search was conducted using the ML parameters identified by
17 Modeltest; Modeltest topology as starting tree and NNI branch swapping. The
18 topology found at the end of the search was used to identify new parameters. These
19 new parameters and topology were used to conduct a new search till convergence.
20 The best ML parameters found were then used to compute 500 bootstrap replicates
21 starting with a NJ tree and setting the maximum number of tree to one to reduce
22 computation time.

23 The phylogenetic tree obtained in this study was in agreement with the results
24 of other studies that divided cyanobacteria into seven or eight major lineages (8, 10),
25 except that group 2 (8) was here paraphyletic (Fig. 1). Sponge-associated

1 cyanobacteria were found to be polyphyletic. They were divided into thirteen lineages
2 spread among various groups of cyanobacteria (Fig. 1 and 2). The major clade of
3 sponge-associated cyanobacteria (37 strains from 18 different sponge species
4 collected from a wide range of geographic regions: Australia, Caribbean, French and
5 Italian Mediterranean coast, Red Sea, Zanzibar) was strongly affiliated with group 6
6 (*Prochlorococcus*, *Synechococcus* and *Microcystis*, (8)). Three additional sponge-
7 associated cyanobacteria (from *Chondrilla* spp. and *Haliclona* sp.) were also part of
8 group 6, but were closer to free-living *Synechococcus* than to the other sponge
9 symbionts (Fig. 2). A second group of sponge-associated cyanobacteria (consisting of
10 four sequences from *Dysidea herbacea*, *Dysidea granulosa*, *Lendenfeldia dendyi* and
11 *Aplysina gerardogreeni*, originating from Guam, Mexican Pacific and Zanzibar) were
12 affiliated to group 2 (morphologically classified in the order Oscillatoriales). A few
13 other sponge-associated cyanobacteria were affiliated with group 5 (Chroococcales,
14 Oscillatoriales, Pleurocapsales and Prochlorales, (8)), but they did not form a
15 monophyletic group (Fig. 1). One additional cyanobacterium (AF295635) from an
16 Australian sponge was found affiliated to the marine group 7 (8). Finally, a sequence
17 of a cyanobacterium associated with the sponge *Pseudoaxinella flava* appeared
18 unrelated to any major group of cyanobacteria (Fig. 1). The polyphyletic origin of
19 sponge-associated cyanobacteria indicates that they derived from multiple
20 independent symbiotic events, involving several different cyanobacterial types and/or
21 host sponges.

22 The phylogenetic relationships between the cyanobacterial sequences of group
23 6 suggest only a partial specificity of cyanobacteria to the host sponges. Two
24 specimens of *Petrosia ficiformis*, one collected in Italy (this study), and another in
25 France (13), were found to be 99% identical, and closely related (BP = 100%). Three

1 cyanobacterial types from two Bahamian *Xestospongia muta* specimens were strongly
2 affiliated to each other and with that from the congeneric species *Xestospongia*
3 *proxima* (BP= 100%). Cyanobacteria from two congeneric *Theonella* species
4 (*swinhoei* and *conica*) were affiliated to each other (BP=77%). Finally, six
5 cyanobacteria associated with *Chondrilla australiensis* (13, 15) were closely affiliated
6 (BP = 49%). These results may also support the idea of co-evolution between sponges
7 and their symbionts. However, cyanobacteria associated to *Theonella swinhoei* from
8 Japan and the Red Sea clustered in different groups (Fig. 1 and 2). In this large-scale
9 phylogenetic analysis, sponge-associated cyanobacteria clades deriving from distant
10 geographic location (as the clades in group 2 and 6) are most probably true-
11 symbionts, while the symbiotic nature of sponge cyanobacteria that are closely related
12 to free living species (*e.g.* those in *Pseudoaxinella flava*) remains uncertain.

13 In the future it will be interesting to examine whether cyanobacterial sponge
14 symbionts from different lineages perform diverse functions in this symbiosis. In
15 addition, reconstruction of sponge molecular phylogeny will enable to test for co-
16 evolution of cyanobacteria and their host sponges.

17

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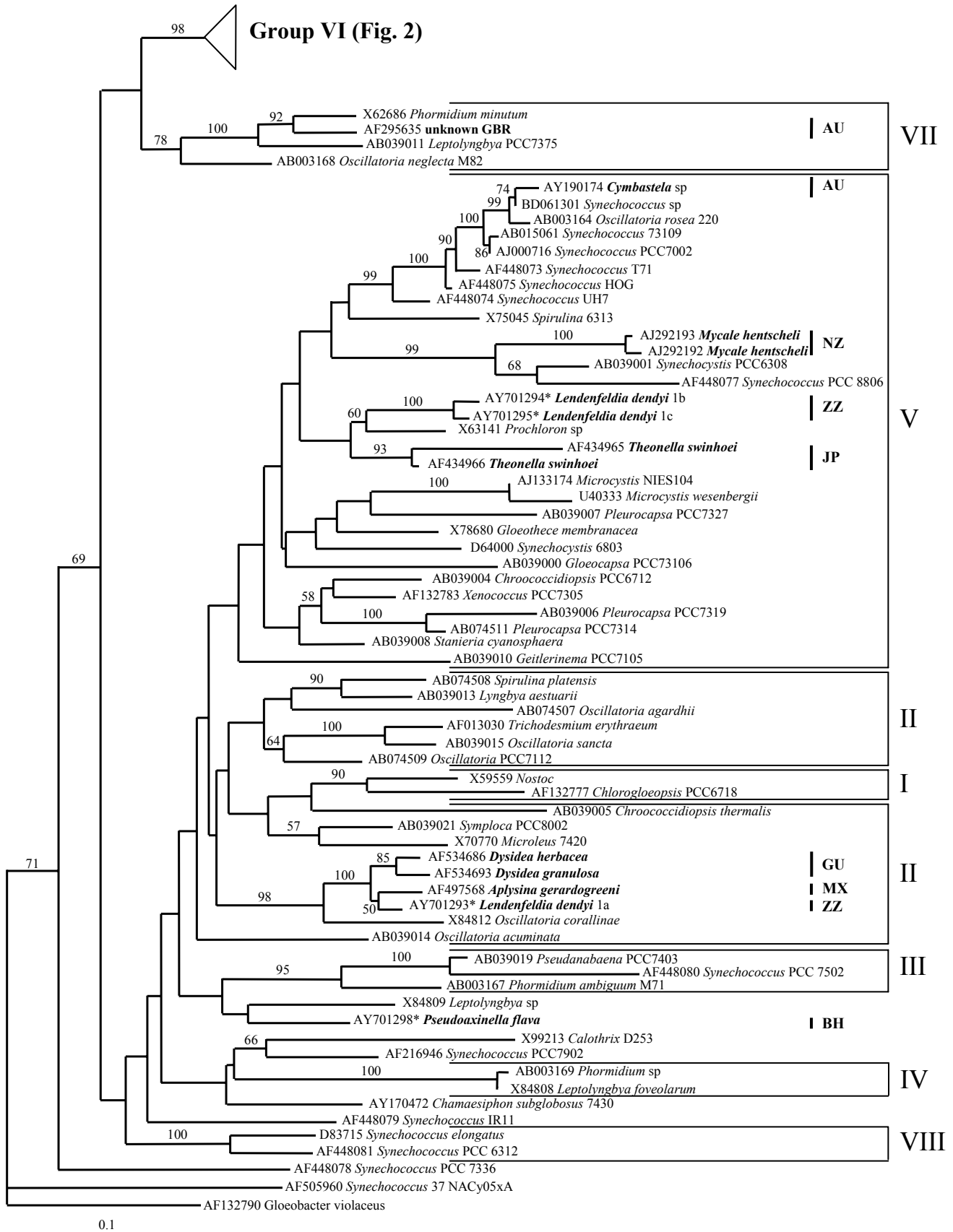
1 FIGURE LEGENDS

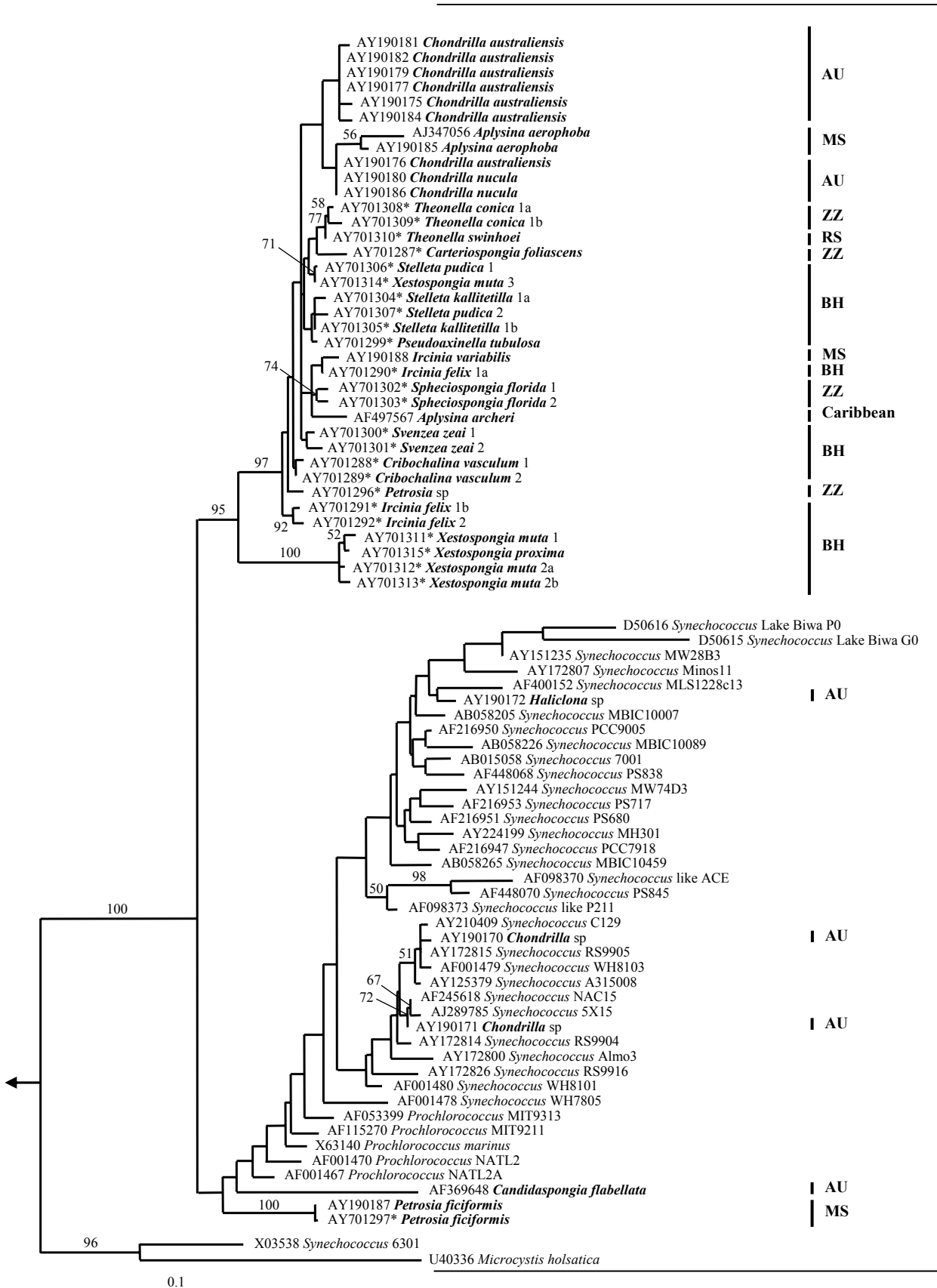
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3 FIG. 1: Maximum Likelihood tree of sponge-associated cyanobacteria based
4 on 16S rRNA. Bootstrap values above 50% are indicated. Sponge-associated
5 cyanobacteria (which were named following their host species) are indicated in bold.
6 The group numbers follow Honda *et al.* (8) and Robertson *et al.* (10). The triangle
7 represents group 6 detailed in Figure 2. The individual identification number is given
8 at the end of the sequence; different letters represent different clones obtained for the
9 same sponge individual. Sequences from this work are indicated by a star after the
10 accession number. The sampling localities of sponge-associated cyanobacteria are
11 indicated by bars. AU-Australia, BH-Bahamas, GU-Guam, JP-Japan, MX-Mexican
12 Pacific, NZ-New Zealand, ZZ-Zanzibar.

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14 FIG. 2: “Group 6” detailed 16S rRNA-based Maximum Likelihood tree of
15 sponge-associated cyanobacteria. Arrow, to outgroup (cf. Fig. 1). Bootstrap values
16 above 50% are indicated. Sponge-associated cyanobacteria (which were named
17 following their host species) are indicated in bold. The individual identification
18 number is given at the end of the sequence; different letters represent different clones
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20 star after the accession number. The sampling localities of sponge-associated
21 cyanobacteria are indicated by bars. AU-Australia, BH-Bahamas, MS-Mediterranean
22 Sea, RS-Red Sea, ZZ-Zanzibar.





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